TOTAL SYNTHESES OF CHIRAL sn-myo-INOSITOL-1,4,5-TRISPHOSPHATE¹ AND ITS ENANTIOMER.

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sn-myo-Inositol-1,4,5-trisphophate (Ins(1,4,5) P_3) and its enantiomers are prepared by synthesis of suitably protected myo-inositols, separation of enantiomers via the formation of D-mannose diastereomeric derivatives and selective phosphorylations.

sn-myo-Inositol-1,4,5-trisphosphate Ins $(1,4,5)P_3$ (-)-8 is a well known second messenger in living cells.² To date, only chemical approaches could provide preparative quantities of inositol-phosphates for physico-chemical and biological investigations. Biological probes, labelled and modified inositol-phosphates can also be obtained only by synthetic means. During the last three years, several syntheses of chiral Ins $(1,4,5)P_3$ have been reported. The separation of the enantiomers was achieved by the formation of diastereomeric esters using chiral acids such as l-menthoxyacetic acid³ or camphanic acid⁴ or by the use of glycosyl derivatives.^{5,6} Some separations of enantiomers were also performed by chiral HPLC methods.⁷ More recently a method using an enzymatic reaction has been proposed.⁶ The aim of our program is to develop convenient methods for the total synthesis of Ins $(1,4,5)P_3$ and derivatives

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which would subsequently be used for biological studies or to study their complexation properties towards cations of biological interest. We want to report here a new synthesis of $Ins(1,4,5)P_3$ enantiomers.

For our synthesis, we used 1(3), 4(6)-di-D-benzyl-2, 3(1)-isopropyliden- sn-myo - inositol (±)-1) as starting material. The synthesis of this compound has been reported by Gigg.⁹ The free hydroxyl groups of diol (±)-1) were then phosphorylated using dianilidophosphoryl chloride.^{10,11} (8 equ, pyridine, -10°C, then 48 h RT, yield 96 % after chromatography, recrystallization from ethanol m.p. 110-111°C). This method gave an excellent yield in the phosphorylation of the vicinal 4,5-diol (±)-2. The next step of the synthesis involved the selective removal of the ketal group⁹, leading to the 1(3),2-dihydroxy derivative (±)-3. Optical resolution of (±)-3 was achieved by means of the formation of diastereomers using a conveniently protected D-mannose. Thus the reaction of 3,4,6-tri-0-acetyl-1,2,0-ethylorthoacetyl -β-D-mannopyranose¹² (D-mannose ethylorthoacetate) 4 with compound (±)-3 under standard conditions¹³ resulted in esterification the enantiotopic positions 1 and 3 of the diol (±)-3 and yielded a mixture of the diastereomers 5 and 6 (Scheme 1).



Scheme 1

These isomers were separated by preparative HPLC to give individual diastereomers¹⁴ (Lichrospher 10 μ M 150x15, acetonitrile/water/triethylamine : 1500/500/1, 9.9 ml. min⁻¹).

Chiral $Ins(1,4,5)P_3$ (-)-8 was obtained from compound 5 using the following steps (Scheme 2). The D-mannose moiety was removed from diastereomer 5 under mild acidic conditions¹³ to yield 4,5-bisdianilidophosphate-3,6-di-0-benzyl *sn-myo*-inositol (+)-3 (82 % recrystallization from ethanol m.p. 181°C, $[\alpha]_p = +34 \pm 1 \text{ c=}2.3$ in chloroform).



Scheme 2

Phosphorylation of (+)-3 again using dianilidophosphoryl chloride leads to the selective phosphorylation of the equatorial hydroxyl group and gave the protected $Ins(1,4,5)P_3$ (+)-7 which was purified by column chromatography (silica gel, chloro form/acetone : 50/1, yield 44.5 %, recristallization from ethanol m.p. 230 -231°C, $[\alpha]_{D}$ = +16 ±1 c=3.5 in chloroform). The complete deprotection was achieved in two steps but in a one pot reaction. First, the anilino groups of the phosphates were removed by reaction with isoamyl nitrite in pyridine, acetic acid and acetic anhydride^{13,15} (1/1/1). Then the reaction mixture was evaporated to dryness and the residue was dissolved in a mixture of ethanol and water (6/1). This solution was hydrogenolysed in the presence of palladium. The crude product was purified by HPLC (Versapack C₁₈ 250x4.6, gradient 0-60% methanol in 0.1 M ammonium formate, pH 5.7, 1 ml. min⁻¹). The elution of $Ins(1,4,5)P_3$ (-)-8, as its ammonium salt, was monitored by the use of (³H)-Ins(1,4,5)P₃ (Amersham)(12.1 %, $[\alpha]_D$ = -28 ±3 c=0.37 in water pH 10, lit. $[\alpha]_D$ = -27.4 in water pH 9.5¹⁶, ³¹P-NMR : 3.37 (<u>d</u>), 5.09(<u>d</u>, 5.18(<u>d</u>)). Our sample was compared with an authentic sample of natural $Ins(1,4,5)P_3$ isolated from beef

heart muscle (paper chromatography, HPLC, ion exchange chromatography) and showed the same chromatographic properties.

The enantiomer of compound (-)-8 Ins $(3,5,6)P_3$ was obtained using the same sequence of reactions as for $Ins(1,4,5)P_3$ but starting from diastereomer 6 (9 %, $[\alpha]_{O}$ =+30 ±3 c=0.29 in water pH 10).¹⁷ The use of D-mannose diastereomeric derivatives could be a general method of preparation of chiral inositol-phosphates.

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REFERENCES AND NOTES

- 1. The configuration of chiral and racemic myo-inositol compounds is described in terms of stereospecific nomenclature. See B.A. Klyashchitskii, V.I. Shvets, W.A. Preobrazhensky, Chem. Phys. Lipids, 1969, 3, 393; and V.I. Shvets, R.P. Evstigneeva, Zhur. Org. Khemi, 1972, 8, 1550.
 M.J. Berridge, R.F. Irvine, Nature, 1984, 312, 315.
 Scherking, V. S
- 3. S. Ozaki, Y. Watanabe, T. Ogasawara, J. Kondo, N. Shiotani, H. Nishii, T. Matsuki, Tetrahedron Lett., 1986, 27, 3157.
- 4. J.P. Vacca, S.J. de Solms, J.R. Huff, J. Amer. Chem. Soc., 1987, 109, 3478. 5. A.E. Stepanov, B.A. Klyashchitskii, V.I. Shvets, R.P. Evstigneeva, Bioorg. Khim., 1976, 2, 1627.
- 6. C.B. Reese, J.G. Ward, Tetrahedron Lett., 1987, 28, 2309.
- 7. S. Ozaki, M. Kohno, H. Nakahira, M. Bunya, Y. Watanabe, Chem. Letters, 1988, 77.
- 8. Y.-C. Liu, C.-S. Chen, Tetrahedron Letters, 1989, 1617.
- 9. J. Gigg, R. Gigg, S. Payne, R. Conant, Carbohydrate Res., 1985, 140C1. 10. V.N. Krylova, N.P. Gornaeva, G.F. Okinik, V.I. Shvets, Zhur. Org. Khim, 1980, 16, 315.
- 11. The phosphorylation procedures were tested for the preparation of racemic $Ins(1,4,5)P_3$. See A.E. Stepanov et al., *Bioorgan. Khim.*, 1989, 15, 850.
- 12. A.E. Stepanov, V.I. Shvets, R.P. Evstigneeva, *Zhur. Org. Khim.*, 1977, 13, 1410. 13. V.I. Shvets, B.A. Klyashchitskii, A.E. Stepanov, R.P. Evstigneeva, *Tetrahedron*, 1975, 29, 331.
- 14. Four peaks could be separated by HPLC. The two main peaks correspond to the diastereomers having the oxo configuration for the inositol residue on the mannose orthoester. The two small peaks correspond to the diastereomers having the endo configuration. ¹H-NMR analysis are in agrement with the expected structures.
- 15. E. Ohtsuka, K. Murao, M. Ubasawa, M. Ikchara, J. Amer. Chem. Soc., 1979, 92, 3441.
 - L.A. Slotin, Synthesis, 1977, 737.
- 16. C. Grado, C.E. Ballou, J. Biol. Chem., 1961, 236, 54. 17. All compounds gave satisfactory C,H,N,P analyses. TLC, IR, UV, ¹H-NMR, ³¹P-NMR, spectra are in aggrement with the expected structures.

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